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THE STATE OF SCIENTIFIC INFORMATION RELATING TO BIOLOGICAL AND ECOLOGICAL
PROCESSES IN THE REGION OF THE GORDA RIDGE, NORTHEAST PACIFIC OCEAN:
BENTHOS

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TABLE OF CONTENTS

	Page
INTRODUCTION	1
GORDA RIDGE LEASE AREA	2
RELATED STUDIES IN THE NORTH PACIFIC	5
HYDROTHERMAL VENTS	9
DATA GAPS	16
ACKNOWLEDGEMENT	18
BIBLIOGRAPHY	20
APPENDIX 1. Species found on the Gorda Ridge or within the lease area	36
APPENDIX 2. Species found outside the lease area that may occur in the Gorda Ridge Lease area, including hydrothermal vent organisms	55

BENTHOS

THE STATE OF SCIENTIFIC INFORMATION RELATING TO THE BIOLOGY AND ECOLOGY OF THE GORDA RIDGE STUDY AREA, NORTHEAST PACIFIC OCEAN:

BENTHOS

INTRODUCTION

Presently, only two published studies discuss the ecology of benthic animals on the Gorda Ridge. Fowler and Kulm (1970), in a predominantly geological study, used the presence of sublittoral and planktonic foraminiferans as an indication of uplift of the deep-sea floor. Their results showed that sediments and foraminiferans are deposited in the Escanaba Trough, in the southern part of the Gorda Ridge, by turbidity currents with a continental origin. They list 22 species of foraminiferans from the Gorda Rise (See Appendix 1). A more recent study collected geophysical, geological, and biological data from the Gorda Ridge, with particular emphasis on the northern part of the Ridge (Clague et al. 1984). Geological data suggest the presence of widespread low-temperature hydrothermal activity along the axis of the northern two-thirds of the Gorda Ridge. However, the relative age of these vents, their present activity and presence of sulfide deposits are currently unknown. The biological data, again with an emphasis on foraminiferans, indicate relatively high species diversity and high density, perhaps associated with widespread hydrothermal activity. Using rock dredges and small sediment collectors, Clague et al. (1984) also collected many sponge spicules, anemones of the genus Actinauge, black corals of the Order Antipatharia, worm tubes, unidentified polychaetes, an unidentified stalked crinoid from the family Hyocrinidae, and the brittle star Dytaster gilberti. Some of the other animals collected include an octopod, pelecypod fragments and an inarticulate brachiopod. These are not animals specifically associated with hydrothermal vents but the overall high abundance of the fauna indicated an abundance of food that may be vent-related. More importantly, the high abundance of agglutinating benthic foraminiferans in muds with the chemical evidence of hydrothermal activity appears to reflect proximity to hydrothermal vents (Clague et al. 1984).

The vast majority of the research in the Gorda Ridge Lease Area was performed off the Newport transect line (an east-west transect extending to approximately 128° W longitude) at the northernmost region of the lease area. Much of this research was done at Oregon State University and all the Atomic Energy Commission studies and theses from OSU related to benthic ecology on Cascadia Abyssal Plain are included in this section. The only taxonomic groups that have been studied in any detail are the polychaetes (Hancock 1969; Fauchald and Hancock 1981), the amphipods (Dickinson 1976; Dickinson and Carey 1978) and the Echinodermata (Carey 1972a; Kyte 1971; McCauley 1967), with particular emphasis on the holothurians (Carney and Carey 1976; Carney 1977; Carey and Carney 1982). A few other studies review the depth distribution of all major invertebrate groups and these are important sources for the species list found in Appendix 1 (Carey and Alsopach 1966; McCauley 1972; Pereyra and Alton 1972).

BENTHOS

There is a paucity of data concerning the benthos of the Gorda Ridge, with most of the research occurring in the northern region of the lease area. Only a few animal groups are adequately covered and most of the research is descriptive, with emphasis on taxonomy and community structure. Little or no data are available on community processes, on the behavior of benthic organisms, and particularly on the smaller meiofauna and microfauna present in the sediments.

GORDA RIDGE LEASE AREA

Taxonomy and Systematics

The papers listed below discuss the taxonomy of the various invertebrate groups and some evolutionary aspects. There is little mention of distribution, abundance or community structure. The majority of this work in the Gorda Ridge lease area has been done off the Newport transect line (44° 39' N) by researchers from Oregon State University. Only the class Polychaeta (Fauchald and Hancock 1981; Hancock 1969) and the phylum Echinodermata (Kyte 1971; McCauley 1967; McCauley and Carey 1967) are covered adequately for this geographic area.

- a. Phylum Annelida
 Class Polychaeta: Fauchald and Hancock 1981
 Hancock 1969
- b. Phylum Arthropoda
 Subphylum Crustacea
 Class Eumalacostraca: Ambler 1980
- c. Phylum Mollusca
 Class Bivalvia: Bernard 1974
- d. Phylum Echinodermata
 Class Ophiuroidea: Kyte 1971
 Class Echinoidea: McCauley and Carey 1967

Community Structure

The diversity (Stander 1970), depth of occurrence (Alton 1972) and species composition of macrofaunal invertebrates (Carey 1966; McCauley 1972) of many phyla are discussed in the general papers. The general papers are sources for the species list found in the Appendices and the research effort is concentrated in the northernmost part of the lease area off the Newport transect line. A more specific report (Carney and Carey 1982), indicated that the diversity of holothurians is highest at a lower bathyal-abyssal depth of 3500 m and catch size decreases with depth. However all other taxonomic groups are poorly described. Data on community processes, such as oxygen consumption, also are lacking for the lease area.

BENTHOS

- a. General: Alton 1972
Carey and Alspach 1966
McCauley 1972
Stander 1970
- b. Specific Taxonomic Groups
 - i. Phylum Echinodermata
Class Holothuroidea: Carney and Carey 1982

Distribution

Pereyra and Alton (1972) is one of the best references for data on the depth distribution of the major epifaunal organisms (particularly the echinoderms) on the abyssal plains north of the Gorda Ridge. More specific papers discuss the depth distribution of amphipods (Dickinson 1976; Dickinson and Carey 1978), of tanner crabs (Pereyra 1966), of scaphopods (Bruce and Carey 1969) and of holothurians (Carney and Carey 1976; Hufford 1967). All these specific studies were conducted off the Newport transect line on Cascadia and Tufts Abyssal Plains. These studies have defined four bathymetric provinces off the coast of Oregon, two of which are in the lease area at depths of more than 2000 m (Carney and Carey 1976). The gammarid amphipod assemblages at a nearshore and an offshore station both at 2800 m depth on Cascadia Plain are very different (Dickinson and Carey 1978). Data on most phyla are lacking for the Gorda Ridge itself and the surrounding areas.

- a. General: Pereyra and Alton 1972
- b. Specific Taxonomic Groups
 - i. Phylum Arthropoda
Subphylum Crustacea
Class Eumalacostraca: Dickinson 1976
Dickinson and Carey 1978
Pereyra 1966
 - ii. Phylum Mollusca
Class Scaphopoda: Bruce and Carey 1969
 - iii. Phylum Echinodermata
Class Holothuroidea: Carney and Carey 1976

Abundance

The studies that discuss the abundance of macrofaunal invertebrates are once again concentrated on the Newport transect line. Carey (1981) sampled an east-west transect across Cascadia and Tufts Abyssal Plains. The biomass of macro-infauna ranges from a low of 1 g wet wt/m² on Eastern Tufts Plain to a high of 8 g wet wt/m² at the base of the continental slope. The numerical density ranged from a low of 200-500 animals/m² on the abyssal plains to a high of 1500 animals/m² at the slope base. In all the samples,

BENTHOS

the polychaetes comprise at least 60% and up to 89% of the animals collected, with the arthropods, mainly crustaceans, representing another 2-25%. Carey and Stander (1968) discuss the technique of using bottom photography for estimating macrofaunal abundance while Carney (1976) found that the abundance of holothurians on Cascadia and Tufts Abyssal Plains is related to depth. Data on other phyla, as well as areas closer to the Gorda Ridge, are lacking.

- a. General: Carey 1981
Carey and Stander 1968
- b. Specific Taxonomic Groups
 - i. Phylum Echinodermata
Class Holothuroidea: Carney 1976

Feeding Ecology

The only group studied in detail are the asteroids. Carey (1972a) and Carey et al. (1967) found that predators decrease in relative abundance from 67% to 0%, deposit feeders decrease from 33% to 14%, and the omnivores increase from 0% to 71% with increasing depth from shallow waters to the abyssal plains. The general conclusion is that deep-sea asteroids are facultative feeders and obtain their food from the sediments, from live prey, and from animal remains. This research was conducted off the Newport transect line. No other data on the feeding ecology of macrofaunal invertebrates has been found for the lease area.

- a. Phylum Echinodermata
 - i. Class Asteroidea: Carey 1972a
Carey et al. 1967

Animal-Sediment Interactions

Studies of animal-sediment interactions all indicate that the organic content and the particle size of the sediment are very important in determining the abundance of infauna (Carey 1965; Carey and Paul 1968). Griggs et al. (1969) studied these interactions on Cascadia Abyssal Plain and particularly in Cascadia Channel and they found that the total number of macroinfauna was four times greater in the channel than on the abyssal plain, probably due to the increased organic content and smaller particle size of the sediments in the channel. No specific taxonomic group has been studied in the lease area.

- a. General: Carey 1965
Carey and Paul 1968
Griggs et al. 1969

BENTHOS

Physiological and Biochemical Processes

Contrary to the rich data on the physiology and biochemistry of vent-associated animals, no data are available on these processes for any phylum in the lease area.

Radioecological Studies

As a part of an intense effort by the U.S. Atomic Energy Commission to determine the effects of radioactive waste off the Columbia River estuary, many studies were done off the Newport transect line to determine the concentration of radionuclides in animals. All the benthic studies (Carey 1963, 1969, 1972c; Carey and Cutshall 1973) found that zinc⁶⁵ specific activity decreases with distance from shore and with depth for all animals investigated, particularly for echinoderms (Carey 1969). Zinc⁶⁵ activity was detected at a depth of 2800 m on Cascadia Abyssal Plain and it was suggested that the organisms absorb this radionuclide through the food web. Again the only data available are from the Newport transect line in the northernmost area of the Gorda Ridge study area.

- a. General: Carey 1963
Carey 1972c
Carey and Cutshall 1973
- b. Specific Taxonomic groups
Phylum Echinodermata
Class Asteroidea: Carey 1969

General Scope

These studies review important physical, chemical, and biological data from cruises off the Columbia River estuary. The data refer to the effect of the Columbia River effluent. One important geological study, Griggs (1969), was included because it studied the sedimentology of Cascadia Channel which is related to other benthic studies on Cascadia and Tufts Abyssal Plains.

- a. Oceanographic data: Anonymous 1966, Vol. I, II, III.
Fleming et al. 1967
- b. Geological: Griggs 1969

RELATED STUDIES IN THE NORTH PACIFIC

Some studies outside the lease area are included in this review because they discuss important deep-sea processes and animals that most likely also occur in the lease area. The taxonomic papers review deep-sea animals found off Washington or British Columbia (Dunn 1982; Banse 1979; Dickinson 1983;

BENTHOS

Johnson et al. 1974; Alton 1966; Lambert 1984). All species mentioned in these papers can be found in Appendix 2.

The most important studies in this section are those of Smith et al. (1979), Smith et al. (1983), and Smith and Baldwin (1984a) that report values of benthic community oxygen consumption in the central and eastern North Pacific Ocean; Burnett (1977) and Snider et al. (1984) that discuss the nannobiota and meiofauna of deep-sea sediments in the central North Pacific; and those of Ingram and Hessler (1983) and Smith and Baldwin (1984b) that studied the distributional and behavioral ecology of scavenging amphipods in the deep-sea below the central North Pacific gyre.

Many other studies on deep-sea benthic ecology could have been included but only those mentioned here have any relevance to the Corda Ridge environment. The relevance of these studies can also be questioned because they are related to deep-sea processes and animals of abyssal plains rather than tectonically active ridges. Nevertheless, the processes they describe may occur in very similar ways in the lease area.

Taxonomy and Systematics

Only a few invertebrate groups are represented in these papers which deal mainly with the taxonomy and evolutionary position of the animals. Dunn (1982) studied the deep-sea anemone Paraphelliactis pabista off British Columbia. Dickinson (1983) worked on the systematics and distribution of the ampeliscid amphipod genera Byblis and Haploops off British Columbia and Lambert (1984) surveyed the deep-sea holothurians off British Columbia. Banse (1979) reviewed the ampharetid polychaetes off British Columbia and the coast of Washington. Alton (1966) described the asteroid species Asthenactis fisheri off the coast of Washington. Johnson et al. (1974) used genetic techniques to differentiate five species of pandalid shrimps from the deep sea off Washington.

- a. Phylum Coelenterata
Class Anthozoa: Dunn 1982
- b. Phylum Annelida
Class Polychaeta: Banse 1979
- c. Phylum Arthropoda
Subphylum Crustacea
Class Eumalacostraca: Dickinson 1983
Johnson et al. 1974
- d. Phylum Echinodermata
Class Asteroidea: Alton 1966
Class Holothuroidea: Lamber 1984

Community Dynamics

Papers that deal with community dynamics outside the lease area are those of Smith et al. (1979), Smith et al. (1983), and Smith and Baldwin (1984a). This research was conducted with a free-vehicle grab respirometer. The study of community oxygen consumption and nutrient exchange was done on an east-west transect from San Diego to Hawaii, south of the lease area but at deep-sea depths in the central and eastern North Pacific. Smith et al. (1979) found no difference in community oxygen consumption between two depths (1193 m and 3815 m) but differences in nutrient exchange between sediments and overlying water. At 1193 m, ammonia, nitrite and phosphate are released to the water and nitrate is taken up by the sediments. At 3815 m, all nitrogenous compounds are evolved and phosphate is taken up. Smith et al. (1983) found that community oxygen consumption, macrofaunal abundance and biomass, and surface sediment organic carbon and total nitrogen all increase along a gradient of increasing primary production from west to east along the transect. Nitrogenous nutrient exchange rates are highly variable and show no east-west trends. Smith and Baldwin (1984a) reported, for the first time, seasonality in biological rates in the deep sea. At both abyssal stations samples, community oxygen consumption is highest in early summer and decreases to the lowest rates in late autumn and winter. No other data that might be important for community processes in the lease area have been found.

Distribution

Two general papers discuss quantitative sampling, distribution and biomass estimates of nannobiota, microbiota, and meiofauna in the deep-sea central North Pacific. Burnett (1977) found that the microbiota of the abyssal central North Pacific consists of ciliates, flagellates, amoebae, testate cells, large prokaryotes, and yeast-like cells at abundances from 16,500 to 26,900 cm^{-2} . The majority of these organisms are restricted to a surface layer about 5 mm thick. Snider et al. (1984) found nannobiota and meiofauna as deep as 5.5 cm in the sediments, but 90% of the numbers and biomass are in the top 3 cm. Foraminifera comprised 86.6% of the meiofaunal biomass.

Another important taxonomic group studied is the scavenging amphipods, particularly Eurythenes gryllus (Ingram and Hessler 1983; Smith and Baldwin 1984b). Again, this research is in the central North Pacific, to the south and west of the lease area. Ingram and Hessler (1983) discovered two groups of scavenging amphipods: a pelagic group of smaller amphipods comprised of Paralicella caperesca, Paralicella tenuipes, and Orchomene gerulicorbis and a demersal group of larger amphipods with Eurythenes gryllus as the only component. All these amphipods are known to exploit food falls and to use chemoreception and possibly mechanoreception to find their food. Smith and Baldwin (1984b) caught Eurythenes gryllus up to 1400 m above the bottom in the deeper, more oligotrophic stations of the central gyre. Maximum catch rates were at 50 m or less above the bottom and larger amphipods were caught at higher altitudes above the bottom. These scavenging amphipods may be equally abundant and important in the Gorda Ridge lease area. Little else

BENTHOS

is known about the distribution of other taxa in areas close to the lease area.

- a. General: Burnett 1977
Snider et al. 1984
- b. Specific Taxonomic Groups
Phylum Arthropoda
Subphylum Crustacea
Class Eumalacostraca: Ingram and Hessler 1983
Smith and Baldwin 1984b

Abundance

Aside from the papers already mentioned in the distribution section (Burnett 1977; Snider et al. 1984), there are no specific papers that pertain to the abundance of important taxonomic groups in areas close to the Gorda Ridge.

Feeding Ecology

There are no specific papers dealing with the feeding ecology of important taxonomic groups in areas closely related to the lease area.

Physiological and Biochemical Processes

There are no data on these processes in areas closely related to the Gorda Ridge lease area.

Radioecological Studies

Carey et al. (1973) studied the effects of munition dumping on benthic invertebrate abundance and composition on the Nitinat deep-sea fan off the coast of Washington, north of the lease area. There were no general trends in faunal composition or abundance that could be directly attributed to the dumping but their conclusions were not definite because no pre-dumping studies had been undertaken. There were some indications of local environmental changes in heavy metal content of the sediments and some faunal changes in the debris area. The work by Cross (1966, 1968), using zinc⁶⁵ as a radioactive metabolic tracer, showed that the shallow-water amphipod Anonyx spp. absorbs zinc⁶⁵ from its most affected when the radionuclide is in high concentration in the sediments. There is also a temperature dependence in the zinc⁶⁵ accumulation and elimination rates.

- a. General: Carey et al. 1973
- b. Specific Taxonomic Groups
Phylum Arthropoda

BENTHOS

Subphylum Crustacea

Class Eumalacostraca: Cross 1966, 1968

HYDROTHERMAL VENTS

Although the presence of hydrothermal vents has yet to be confirmed on the Gorda Ridge, data gathered on the NOAA cruise of April/May 1985 has revealed some radon²³² and particularly some manganese anomalies of the type often associated with hydrothermal activities (R. Collier pers. comm.). These manganese anomalies in the northern part of the Gorda Ridge, in the Narrow Gate region, coincide with areas of tectonic and volcanic activity on the ridge (L.D. Bibee pers. comm.). Also, recent studies on the subduction zone off the coast of Oregon and Washington have revealed the presence of vent-type communities (Suess et al., in press; Kulm et al., unpub. ms.). The biological communities that have been discovered include a vesicomid clam that is probably Calyptogena magnifica, a gutless clam of the genus Solemya and a vestimentiferan worm, Lamellibrachia barhami. Thus the preliminary information suggests that there may be hydrothermal vents on the Gorda Ridge and that there are vent/seep biological communities in the lease area, at least in the northern region off Newport. This means that the research done at other hydrothermal sites, such as the Galapagos Rift, 21°N and 13°N on the East Pacific Rise, and the Juan de Fuca Ridge, will be useful in understanding the biology and physiology of vent-associated organisms.

The hydrothermal vent references included in this section reflect research conducted for the most part in the last 10 years. The studies can be categorized in three ways: (1) systematic and taxonomic studies of the macrofaunal organisms, such as the pogonophoran worm Riftia pachyptila (Jones 1980, 1981, 1984), the large white clam Calyptogena magnifica (Boss and Turner 1980), the galatheid and brachyuran crabs around the vents (Williams 1980; Williams and Chase 1982; Williams and Van Dover 1983) and the chemoautotrophic bacteria present in hydrothermal waters (Baross et al. 1980; Jannasch and Wirsen 1981; Jannasch 1984); (2) descriptive work discussing the community structure of vent-associated organisms (Grassle 1982, 1984a, 1984b; Hessler and Smithey 1984; Lonsdale 1977; (3) physiological and biochemical studies concerning the vent organisms' adaptations to pressure, temperature, toxic chemicals (sulfides), and the chemosynthetic food web of the vents (Arp and Childress 1981a, 1981b; Mickel 1982; Somero 1984; Felbeck 1985).

Taxonomy and Systematics

Much of the research on hydrothermal vent organisms is focussed on the taxonomy and description of the animals associated with the vents. The papers consist of work on all vents from the Eastern Pacific, i.e., Galapagos, 13°N, and 21°N, with the new volume edited by M.L. Jones (in press) also including the biology of vent animals from the Juan de Fuca Ridge. The main taxonomic groups that have been analyzed are the bacteria (Baross et al. 1980; Harwood et al. 1982; Jannasch and Wirsen 1981 and

BENTHOS

others listed below), the polychaetes (Desbruyeres and Laubier 1980, 1982; Fauchald 1982; Maciolek 1981 and others listed below), the pogonophorans, particularly Riftia pachyptila (Jones 1980, 1981, 1984), the crustaceans, particularly the Eumalacostracans (Hessler 1984; Williams 1980 and others), and the molluscs (Boss and Turner 1980; Fretter et al. 1981; Kenk and Wilson in press). The clam Calyptogena magnifica is associated with hydrothermal activity and is one of the largest vesicomid clams, while the gastropod Neomphalus fretterae is not related to any living prosobranch and may represent an important link in gastropod evolution. A few other taxa have been investigated but much more research is needed to identify all vent related organisms.

- a. Bacteria: Baross et al. 1980
Harwood et al. 1982
Jannasch and Wirsén 1981
Jones et al. 1983
Ruby et al. 1981
Stahl et al. 1984
- b. Phylum Coelenterata
Class Anthozoa: unknown
- c. Phylum Annelida
Class Hirudinea: Burreson 1981
Class Polychaeta: Desbruyeres and Laubier 1980, 1982
Fauchald 1982
Haymon et al. 1984
Maciolek 1981
Pettibone 1983, 1984, 1985
Zottoli 1983
- d. Phylum Pogonophora: Jones 1980, 1981, 1984
Tunnicliffe and Juniper 1983
- e. Phylum Arthropoda
Subphylum Chelicerata
Class Acarina: Krantz 1982
Subphylum Crustacea
Class Cirripedia: Newman 1979
Class Copepoda: Humes and Dojiri 1980
Class Eumalacostraca: Hessler 1984
Williams 1980
Williams and Chase 1982
Williams and Van Dover 1983
- f. Phylum Mollusca
Class Bivalvia: Boss and Turner 1980
Kenk and Wilson in press
Class Gastropoda: Fretter et al. 1981
McLean 1981
- g. Phylum Hemichordata
Class Enteropneusta: Woodwick and Sensenbaugh 1985

BENTHOS

h. Phylum Chordata

Class Osteichthyes: Cohen and Haedrich 1983

Community Dynamics

Much of the information on hydrothermal vents concerns the community structure of the organisms. Some of the more general papers discuss the discovery of unusual communities of animals in the deep-sea associated with hydrothermal fluid venting and their relation to a chemosynthetic food source (Corliss et al. 1979; Desbruyeres et al. 1982; Enright et al. 1981; Galapagos Biology Expedition Participants 1979; Grassle 1982). Lonsdale (1977) reported the high standing crop of suspension feeding macrobenthos near hydrothermal vents and related this high abundance to the increased food supply near hydrothermal plumes in bottom water. Desbruyeres and Laubier (1984) review all the data on the primary consumers of Eastern Pacific hydrothermal vents, with an emphasis on the four most important species Riftia pachyptila, Calyptogena magnifica, the mytilid mussel Bathymodiolus thermophilus and the polychaete Alvinella pompejana. Paul et al. (1984) discuss the discovery of cold seep communities at the Florida Escarpment. A recent paper by Smith (1985a) reports some in situ rates of oxygen consumption. He reports that diffuse warm-water effluent along crevices in pillow lava formations are densely populated with epibenthic megafauna, that vent and non-vent plankton are dominated by the calanoid copepod, Isaacsicalanus paucisetus, and that weight-specific oxygen consumption rates of plankton from vent and non-vent areas are of similar magnitude ($1.68 - 4.26 \mu O_2$ (mg dry wt)⁻¹d⁻¹). These oxygen consumption rates are one to two orders of magnitude lower than published rates for surface-water plankton.

The specific taxonomic groups covered include the bacteria, the bivalve molluscs, particularly Calyptogena magnifica and Bathymodiolus thermophilus. from the Galapagos Rift vents, and the gastropod Neomphaulus fretterae. The work on bacteria is focused on their growth at high temperatures (Baross and Deming 1983), their activity in high temperature and high sulfur environments (Karl et al. 1984; Tuttle et al. 1983), and their importance as chemolithoautotrophic primary producers at the vents (Baross et al. 1982; Felbeck and Somero 1982; Jannasch 1984a, 1984b; Jannasch and Wirsen 1979; Karl et al. 1980). Finally the potential of hydrothermal vents as a site for the origin of life is reviewed by Corliss et al. (1980).

The more detailed research on the bivalves near hydrothermal vents reviews the growth rates of the mytilid mussel Bathymodiolus thermophilus the vesicomid clam Calyptogena magnifica. As an example, Turekian and Cochran (1981) found that the vesicomid clam from the Galapagos Rift vents grows about 4 cm per year with an age of 3 or 4 years while Rhoads et al. (1981) found that the mussel grows about 1 cm per year with an approximate lifetime of 19 years. Some research has also been done on the larval development and dispersal of bivalves at vents and it is suggested that the mussel may have a planktotrophic stage for better dispersal while the clam Calyptogena magnifica and many other gastropods have a nonplanktotrophic larval stage (Lutz et al. 1980, 1984, in press; Turner and Lutz 1984; Turner

BENTHOS

et al., in press). The research on Neomphalus fretterae by McLean (1981) indicates that this gastropod is close to many archaeogastropods and may be an ultimate expression of a basic plan that occurred only once in evolution. Little data on the community structure of the other vent-associated animals are available but new photographic evidence on Eastern Pacific vents may provide data on other vents and on the crustaceans living in and around the vents (R.R. Hessler pers. comm.).

- a. General: Cohen and Haedrich 1983
 - Corliss et al. 1979
 - Desbruyeres and Laubier 1984
 - Desbruyeres et al. 1982
 - Enright et al 1981
 - Galapagos Biology Exped. Part. 1979
 - Grassle 1982, 1984a and b
 - Hessler 1981
 - Lonsdale 1977
 - Paul et al. 1984
 - Turner 1981
- i. Oxygen Consumption: Smith 1984
- b. Specific Taxonomic Groups
 - i. Bacteria: Baross and Deming 1983
 - Baross et al. 1982
 - Corliss et al. 1980
 - Felbeck and Somero 1982
 - Jannasch 1984a and b
 - Jannasch and Wirsén 1979
 - Karl et al. 1980, 1984
 - Tuttle et al. 1983
 - ii. Phylum Mollusca
 - Class Bivalvia: Fatton and Roux 1981a and b
 - Fatton et al. 1982
 - Lutz et al. 1980; 1984; in press
 - Rhoads et al. 1981; 1982
 - Roux et al. 1982
 - Smith 1985b
 - Turekian et al. 1979; 1983
 - Turekian and Cochran 1981
 - Turner and Lutz 1984
 - Turner et al. in press
 - Class Gastropoda: McLean 1981

BENTHOS

Distribution

General references on the distribution of vent organisms include Corliss et al. (1979), Crane and Ballard (1980), and Hessler and Smithey (1984). In general, animals such as the vestimentiferan, limpets, clams, a shrimp, an anemone and the mussel live in the mouth of the vents, where the temperature is several degrees above ambient. Other organisms like the serpulid polychaetes, a second anemone, a galatheid crab and a turid gastropod are abundant around the vents but avoid the vent openings. Finally, a third group consisting of a siphonophore, a brachiopod, a third species of anemone, enteropneusts, another shrimp, and an ophiuroid remain at the periphery of the vent field. A few other mobile species, such as vent fish, brachyuran crabs, other galatheid crabs, and amphipods, are most abundant at vent openings but are also found in non-vent areas. Most vent field species are endemic. The main food sources are symbiotic chemoautotrophic bacteria, suspended bacteria ejected from the vents, bacteria settled out from the vents, and bacteria growing as a film on the substratum surrounding the vent fields. Little is known of the plankton associated with these vents but the top of the food chain is represented by scavengers, mainly eumalacostracan crustaceans, some of which may be carnivorous.

Abundance

There are no references that discuss the abundance per se of vent-associated organisms. Some of the general papers on community structure give an indication of the relative abundance of Riftia pachyptila and Calyptogenia magnifica at different vent sites (Desbruyeres and Laubier 1984; Grassle 1982, 1984a, 1984b). There are no papers that parallel the ones discussed in this section for the Gorda Ridge lease area or its close surrounding areas.

Feeding Ecology

Though many of the papers on vent organisms discuss feeding mechanisms, the studies were based on physiological, enzymatic, and biochemical results. These will be discussed in the separate section on Physiological and Biochemical Processes. The only paper resembling those discussed for the other areas reviewed is the one by Van Praet (in press) which was unavailable for comment.

Animal-Sediment Interactions

There are no references related to hydrothermal vents that discuss the interaction between benthic animals and the sediments.

BENTHOS

Physiological and Biochemical Processes

One of the major thrusts in research on hydrothermal vent animals has been to study their physiological and biochemical adaptations for life in a high pressure, elevated temperature, and potentially chemically toxic environment. The techniques used include thin-sectioning histological research, transmission electron microscopy, enzymology, and the use of N^{15}/N^{14} and C^{13}/C^{12} ratios to determine the physiology and biochemistry of feeding of vent-associated organisms. The general papers review the evidence for chemoautotrophy in vent bacteria, the use of hydrogen sulfide as an energy source (Fry et al. 1983), and the symbiotic relationship between sulfide-oxidizing bacteria and the vestimentiferan worm and bivalve molluscs living in the vent openings (Felbeck et al. 1981; Hand and Somero 1983; Somero 1984). Rau's work (1981b) suggested that organic nitrogen of nutritional importance to vent animals is initially synthesized within the vent environment with some evidence of N_2 fixation.

The main specific taxonomic groups reviewed are the bacteria (Cavanaugh 1983; Fisher et al. 1983; Ruby and Jannasch 1982 and others), the vestimentiferan Riftia pachyptila (Arp and Childress 1981a, 1983; Felbeck 1981; Nelson et al. 1984 and many others), the brachyuran crab Bythograea thermydron (Arp and Childress 1981b; Mickel 1982; Mickel and Childress 1982), and the bivalves Calyptogena magnifica (Arp et al. 1984; Childress and Mickel 1982; Roesijadi and Crecelius 1984 and others) and Bathymodiolus thermophilus (Rau et al. 1979; Williams et al. 1981). Some of the most important results include: (1) the existence of chemoautotrophic symbiotic bacteria in many of the vent animals (Cavanaugh 1983; Cavanaugh et al. 1981; Felbeck 1981); (2) the genetic similarity, based on RNA sequences, of the prokaryotic symbionts of Riftia pachyptila, Calyptogena magnifica, and a shallow-water bivalve, Solemya velum (Stahl et al. 1984); (3) the blood of the vent vestimentiferan worm and the vent crab Bythograea thermydron has a high oxygen affinity, a small effect of carbon dioxide on oxygen affinity and a high oxygen carrying capacity (Arp and Childress 1981a and b; Terwilliger et al. 1980; Wittenberg et al. 1981); (4) the C^{13}/C^{12} ratios of vent mussels, vent clams and vent vestimentiferans are all depleted relative to the tissues of other marine organisms which suggests a chemosynthetic food source, either by symbiotic arrangement with chemoautotrophic sulfide-oxidizing bacteria or by ingestion of vent bacteria ejected in vent fluids or growing on microbial mats (Rau et al. 1979; Southward et al. 1981; Williams et al. 1981).

A few other taxa have been investigated with results on the presence of chemosynthetic symbiotic bacteria in oligochaetes (Felbeck et al. 1983), in polychaete worms near hydrothermal vents (Desbruyeres et al. 1983; Laubier et al. 1983, in press), and in shallow-water coastal hydrothermal vent gastropods (Stein 1984).

- a. General: Felbeck et al. 1981
- Fry et al. 1983
- Hand and Somero 1983
- Rau 1981b
- Somero 1984

BENTHOS

b. Specific Taxonomic Groups

- i. Bacteria: Baross and Deming 1983
Cavanaugh 1983
Felbeck and Somero 1982
Fischer et al. 1983
Jannasch and Wirsen 1979
Ruby and Jannasch 1982
Stahl et al. 1984
Trent et al. 1984

- ii. Phylum Annelida

- Class Polychaeta: Desbruyeres et al. 1983
Felbeck et al. 1983
Laubier et al. 1983
Laubier et al in press

- iii. Phylum Pogonophora: Arp and Childress 1981a, 1983
Blum and Fridovich 1984
Cavanaugh et al. 1981
Felbeck 1981, 1985
Nelson et al. 1984
Powell and Somero 1983
Rau 1981a
Southward et al. 1981
Terwilliger et al. 1980
Williams et al. 1981
Wittenberg et al. 1981

- iv. Phylum Arthropoda

- Subphylum Crustacea

- Class Eumalacostraca: Arp and Childress 1981b
Mickel 1982
Mickel and Childress 1982

- v. Phylum Mollusca

- Class Bivalvia: Arp et al. 1984
Blum and Fridovich 1984
Childress and Mickel 1982
Felbeck 1983
Rau 1981a
Rau and Hedges 1979
Roesijadi and Crecelius 1984
Terwilliger et al. 1983
Williams et al. 1981

- Class Gastropoda: Stein 1984

BENTHOS

Radioecological Studies

No studies on the effects of radionuclides or heavy metals have been done for animals from hydrothermal vent habitats. Thus there are no references paralleling the radioecological studies done in the northern part of the lease area.

General Scope

These studies review some of the important geological, chemical, and general oceanographic data on hydrothermal vents. Ballard (1977) reviews the general oceanographic data concerning the Galapagos Rift vents while Lonsdale (1984) does the same for the hot vents of the Sea of Cortez. The work by Edmond (1982) and Edmond et al. (1982) discusses the chemistry of Galapagos hydrothermal vents, particularly the deposition of sulfides and the effects of the plume, while Normark et al. (1982) reviews similar data for the Juan de Fuca Ridge. The importance of sinking particulate matter and the advection of organic material near vents was studied by Cobler and Dymond (1980) and Comita et al. (1984).

- a. Oceanographic data: Ballard 1977
Cobler and Diamond 1980
Comita et al. 1984
Edmond 1982
Edmond et al. 1982
Elvers et al. 1974
Lonsdale 1984
Normark et al. 1982

DATA GAPS

Based on the literature reviewed, it is apparent that only limited data exist concerning the benthos of the Gorda Ridge and the immediate surrounding area. There are only two published studies (Fowler and Kulm 1970; Clague et al. 1984) that deal with the benthos of the Gorda Ridge and only the foraminiferans are discussed in detail. Virtually everything else needs to be assessed, including the taxonomy of the infauna (both meiofauna and macrofauna), the potential hydrothermal communities, community structure, biomass, abundance, distribution, trophic relationships, reproduction and response to disturbances. There are also no data on animal-sediment interactions on and close to the Gorda Ridge, nor any information on heavy metal and radionuclide concentrations in potentially important regions of the Gorda Ridge.

Areas within the lease area have been studied in more detail but there are still major gaps in our knowledge of the benthos in the lease area. The region to the north and east of the Ridge, Cascadia Abyssal Plain specifically, has been studied in detail for the major macro- and megafaunal animals. However, data on nannobiota, meiofauna, and important community processes such as oxygen consumption, nutrient fluxes, feeding ecology,

BENTHOS

trophic dynamics, and reproduction are lacking. All other regions adjacent to the Gorda Ridge and within the lease area have been poorly studied.

Finally, even with all the information known about hydrothermal vent communities, much more research needs to be done to understand fully the ecology of these unique assemblages. More research is needed concerning the taxonomy of vent organisms, the overall community structure including data on abundance and distribution, the physiological adaptations to vent environments, geographic extent of these unusual deep-sea habitats, reproductive and dispersal patterns, and the response of the organisms to disturbance.

In summary, little is known about the benthos of the Gorda Ridge and the surrounding region. Research focusing on the areas outlined below will be useful in gaining insight and knowledge about benthic community processes at Gorda Ridge.

Taxonomy and Systematics

Many of the major taxonomic groups have not been studied including: bacteria, anemones and other coelenterates, pogonophoran worms, chelicerates (particularly pycnogonids), crustaceans (particularly copepods, cirripedes, and eumalacostracans), polychaetes, molluscs, and gastropods in general, and nanobiota and meiofaunal-size organisms.

Community Dynamics

Data on the diversity and density of all major phyla are lacking, with the exception of holothurians on Cascadia Abyssal Plain. Community processes such as oxygen consumption and nutrient fluxes have also not been studied in the lease area.

Distribution

The depth distribution of most major phyla has been researched quite extensively on Cascadia Abyssal Plain but data are lacking for the Gorda Ridge and other regions of the lease area. Research is also needed on the horizontal scales of distribution of organisms living in the lease area, particularly for those organisms on the Ridge per se.

Abundance

The only data available discuss the abundance of infaunal animals on Cascadia Plain and of the holothurians on both Cascadia and Tufts Abyssal Plains. All other taxonomic groups are unstudied and no data exist for the other regions of the lease area or for the Ridge itself. Reproductive biology is an especially important process for which no data exist.

BENTHOS

Feeding Ecology

Data are needed on the feeding ecology of all taxa in the lease area. Nothing is known about the feeding of the sessile fauna surrounding the Ridge, nor on the scavenging amphipods that may be present in the area. Again, the only region with any coverage is the Newport transect line, so all other regions of the lease area must be studied.

Animal-Sediment Interactions

No specific taxonomic group has been studied in the lease area. Little is known about the sediments around the Gorda Ridge and less about the animals living in or on them. Research in this field is particularly important due to the possibility of major sediment disturbances on and around the Ridge. Data are needed on the particle size of the sediments and their organic content, on the rates of bioturbation, and on the physical characteristics of the benthic boundary layer, especially the spatial and temporal variations in current speeds.

Physiological and Biochemical Processes

In light of the recent evidence suggesting active hydrothermal venting at the Gorda Ridge, there is the possibility that vent-related benthic communities will be present. If so, research will be needed on the taxonomy, genetics, and physiology of all animals found on the Gorda Ridge. There are some data on the animals found at the subduction zone off Oregon (Suess et al., in press; Kulm et al., unpub. ms.) but this is the only physiological information on vent-related or any other fauna in the lease area.

Radioecological Studies

Again the research is concentrated on the Newport transect line, in the northernmost region of the lease area, and nothing is known for any other regions of the lease area, including the Gorda Ridge. Basic research on heavy metal concentration and radionuclide content of the sediments should be undertaken to provide good background data for future reference.

ACKNOWLEDGEMENT

Many of the references included in this report were gathered through an exhaustive computer search using the following data bases: Aquatic Sciences and Fisheries Abstracts, Biological Abstracts, National Technical Information Service, and National Environmental Data Referral Service. In addition to the computer search, information and references were obtained from the Atomic Energy Commission Reports and from theses from the College of Oceanography at Oregon State University. The Columbia River effluent cruise reports were obtained on loan from the School of Oceanography of the

BENTHOS

University of Washington. Finally, we obtained some other data and references through personal contacts at Tetra Tech Inc., at Scripps Institution of Oceanography, and particularly at Oregon State University.

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Appendix 1

Species found on the Gorda Ridge or within the lease area

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2. McCauley 1972
3. Suess et al. 1985
4. Kulm et al. 1985
5. Pereyra & Alton 1972
6. Carey 1963
7. Fauchald & Hancock 1981
8. Dickinson & Carey 1973
9. Dickinson 1976
10. Carey & Alspach 1966
11. Ambler 1980
12. Bernard 1974
13. Bruce & Carey 1969
14. Carey et al. 1967
15. Carey 1972a
16. Alton 1972
17. McCauley 1967
18. Carney & Carey 1976
19. Carney 1977
20. Carney & Carey 1982
21. Kyte 1971
22. Fowler & Kulm 1970

BENTHOS

Species known to occur on the Gorda Ridge or in the lease area.
 *=collected on the Gorda Ridge.

Phylum Sarcodina

<u>Adercotryma glomeratum</u> *	1
<u>Ammodiscus spp</u> *	1
<u>Ammolagena</u> *	1
<u>Ammomarginulina</u> *	1
<u>Astrorhiza</u> *	1
<u>Bathysiphon</u> *	1
<u>Bolivina decussata</u> *	22
<u>Bottelina</u> *	1
<u>Succella frigida</u> *	22
<u>Bulimina rostrata</u> *	22
<u>Bulmina subacuminata</u> *	
<u>Buliminella elegantissima</u> *	22
<u>Cassidulina minuta</u> *	22
<u>Cassidulina subglobosa</u> *	22
<u>Cassidulina translucens</u> *	22
<u>Cibicides lobatulus</u> *	22
<u>Cribostomoides</u> *	1
<u>Cyclamina</u> *	1
<u>Cystamina</u> *	1
<u>Eggerella</u> *	1
<u>Eilohedra levicula</u> *	22
<u>Elphidium clavatum</u> *	22
<u>Elphidium microgranulosum</u> *	22
<u>Epistominella pacifica</u> *	22
<u>Eponides tumidulus</u> *	22
<u>Eponides spp.</u> *	22
<u>Globigerina bulloides</u> *	22
<u>Globigerina pachyderma</u> *	22
<u>Globigerina quingueloba</u> *	22
<u>Globigerinita glutinata</u> *	22
<u>Globigerinita uvula</u> *	22
<u>Globotextularia anceps</u> *	1
<u>Glomospira</u> *	1
<u>Haplofragmoides columbiense</u> *	1
<u>Hemisphaerammina</u> *	1
<u>Hormosina globulifera</u> *	1
<u>Hyperammina elongata</u> *	1
<u>Karrerella</u> *	1
<u>Lituotuba</u> *	1
<u>Martinotiella</u> *	1

BENTHOS

<u>Protobotellina</u> *	1
<u>Psammosphaera fusca</u> *	1
<u>Psammosphaera parva</u> *	1
<u>Recurvoides contortus</u> *	1
<u>Reophax difflugiformis</u> *	1
<u>Reophax distans</u> *	1
<u>Reophax guttifer</u> *	1
<u>Reophax nodulosus</u> *	1
<u>Rhabdammina abyssorum</u> *	1
<u>Rhabdammina irregularis</u> *	1
<u>Rhizammina algaeformis</u> *	1
<u>Saccammina</u> *	1
<u>Saccorhiza ramosa</u> *	1
<u>Spiroplectammina spp</u> *	1
<u>Textularia</u> *	1
<u>Thurammina</u> *	1
<u>Tolypammina</u> *	1
<u>Trifarina angulosa</u> *	22
<u>Trochammina</u> *	1
<u>Trochamminoides</u> *	1
<u>Uvigerina auberiana</u> *	22
Subfamily Diffusilinidae *	
Phylum Coelenterata	
Class Hydrozoa	
<u>Lictorella cervicornis</u>	2
<u>Lafoea spp</u>	2
<u>Sertularella tanneri</u>	3,4
Class Anthozoa	
<u>Actinauge spp.</u> *	1
<u>Callogorgia spp.</u>	5
<u>Clavularia spp</u>	2
<u>Epizoanthus spp</u>	2
<u>Kophoblemnon spp</u>	2
<u>Parastenella doederleini</u>	3,4
<u>Pennatula spp</u>	2
<u>Protoptilum spp</u>	2
<u>Zoanthidea spp</u>	1
Order Antipatharian *	
Phylum Echiuroidea	
<u>Listriolobus hexamyotus</u>	2,6

BENTHOS

<u>Thalassema</u> <u>steinbecki</u>	2
Phylum Brachiopoda	
<u>Pelagodiscus</u> <u>atlanticus</u>	2
Phylum Annelida	
Class Polychaeta	
Order Amphinomida	
Family Amphinomidae	
<u>Chloeia</u> <u>pinnata</u>	5,7
<u>Paramphinome</u> <u>pacifica</u>	7
Order Capitellida	
Family Arenicolidae	
<u>Arenicola</u> <u>marina</u>	7
<u>Arenicola</u> <u>pusilla</u>	7
<u>Branchiomaldane</u> <u>vincenti</u>	7
Family Capitellidae	
<u>Capitella</u> <u>capitata</u>	7
<u>Dasybranchus</u> <u>glabrus</u>	7
<u>Heteromastus</u> <u>filiformis</u>	7
<u>Mediomastus</u> <u>californiensis</u>	7
<u>Neoheteromastus</u> <u>lineus</u>	7
<u>Notomastus</u> <u>lineatus</u>	7
<u>Notomastus</u> <u>magnus</u>	7
<u>Notomastus</u> <u>precocis</u>	7
<u>Notomastus</u> <u>tenuis</u>	7
Family Maldanidae	
<u>Asychis</u> <u>disparidentata</u>	7
<u>Asychis</u> <u>ramosus</u>	7
<u>Asychis</u> <u>similis</u>	7
<u>Axiothella</u> <u>rubrocincta</u>	7
<u>Clymaldane</u> <u>laevis</u>	7
<u>Euclymene</u> <u>reticulata</u>	7
<u>Maldane</u> <u>cristata</u>	7
<u>Maldane</u> <u>glebifex</u>	7
<u>Maldane</u> <u>harai</u>	5
<u>Maldane</u> <u>monilata</u>	7
<u>Maldane</u> <u>sarsi</u>	5,7
<u>Nicomache</u> <u>lumbricalis</u>	5,7
<u>Notoproctus</u> <u>pacificus</u>	7
<u>Praxillella</u> <u>affinis</u> <u>pacifica</u>	7

BENTHOS

<u>Praxillella gracilis</u>	5,7
<u>Rhodine bitorquata</u>	7
Order Cossurida	
Family Cossuridae	
<u>Cossura brunnea</u>	7
<u>Cossura longicirrata</u>	7
<u>Cossura modica</u>	7
<u>Cossura rostrata</u>	7
Order Eunicea	
Family Arabellidae	
<u>Arabella iricolor</u>	7
<u>Drilonereis falcata</u>	7
Family Dorvilleidae	
<u>Dorvillea bati</u>	7
<u>Dorvillea pseudorubrovittata</u>	7
<u>Protodorvillea gracilis</u>	7
<u>Schistomeringos longicornis</u>	7
Family Eunicidae	
<u>Eunice kobiensis</u>	5,7
<u>Eunice segregata</u>	7
<u>Marphysa stylobranchiata</u>	7
Family Lumbrineridae	
<u>Lumbrineris abyssicola</u>	7
<u>Lumbrineris bricirrata</u>	7
<u>Lumbrineris cruzensis</u>	7
<u>Lumbrineris eugeniae</u>	7
<u>Lumbrineris index</u>	7
<u>Lumbrineris lagunae</u>	7
<u>Lumbrineris latreilli</u>	7
<u>Lumbrineris moorei</u>	7
<u>Lumbrineris sarsi</u>	7
<u>Lumbrineris similabris</u>	7
<u>Lumbrineris zonata</u>	7
<u>Ninoe fusca</u>	7
<u>Ninoe fuscoides</u>	7
<u>Ninoe gemma</u>	7
<u>Ninoe longibranchia</u>	7
Family Onuphidae	
<u>Diopatra ornata</u>	7

BENTHOS

<u>Hyalinoecia stricta</u>	7
<u>Nothria conchylega occidentalis</u>	5,7
<u>Nothria elegans</u>	6,7
<u>Nothria geophiliformis</u>	7
<u>Northria iridescens</u>	5,7
<u>Nothria lepta</u>	5,7
<u>Northria mixta</u>	7
<u>Nothria pallida</u>	7
<u>Nothria stigmatis</u>	7
<u>Onuphis parva</u>	7
<u>Onuphis profundi</u>	7
<u>Onuphis vexillaria</u>	7
Order Flabelligerida	
Family Fauveliopsidae	
<u>Fauveliopsis armata</u>	7
<u>Fauveliopsis glabra</u>	7
<u>Fauveliopsis magna</u>	7
Family Flabelligeridae	
<u>Brada pluribranchiata</u>	7
<u>Brada villosa</u>	7
<u>Diplocirrus micans</u>	7
<u>Flabelligera infundibuliformis</u>	7
<u>Pherusa inflata</u>	7
<u>Pherusa negligens</u>	7
<u>Pherusa papillata</u>	7
<u>Uncopherusa bifida</u>	7
Order Opheliida	
Family Opheliidae	
<u>Armandia brevis</u>	7
<u>Euzonus mucronata</u>	7
<u>Euzonus williamsi</u>	7
<u>Ophelia limacina</u>	7
<u>Ophelina acuminata</u>	7
<u>Ophelina breviata</u>	7
<u>Travisia brevis</u>	5,7
<u>Travisia foetida</u>	7
<u>Travisia gigas</u>	7
<u>Travisia oregonensis</u>	7
<u>Travisia pupa</u>	5,7
Family Scalibregmidae	
<u>Mucibregma spinosa</u>	7
<u>Scalibregma inflatum</u>	5,7

BENTHOS

Order Orbiniida

Family Orbiniidae

<u>Califia mexicana</u>	7
<u>Haploscoloplos elongatus</u>	7
<u>Haploscoloplos kerguelensis</u>	7
<u>Naineris dendricata</u>	7
<u>Naineris uncinata</u>	7
<u>Phylo nudus</u>	7
<u>Scoloplos acmeceps</u>	7
<u>Scoloplos armiger</u>	7
<u>Scoloplos mazatlanensis</u>	7

Family Paraonidae

<u>Acesta lopezi</u>	7
<u>Aedicera antennata</u>	7
<u>Aedicera longicirrata</u>	7
<u>Aedicera oregonensis</u>	7
<u>Aedicera pacifica</u>	7
<u>Allia crassicapitis</u>	7
<u>Allia hartmani</u>	7
<u>Allia ramosa</u>	7
<u>Aricidea suecica</u>	7
<u>Paraonella abranchiata</u>	7
<u>Paraonella cedroensis</u>	7
<u>Tauberia gracilis</u>	7

Order Oweniida

Family Oweniidae

<u>Myriochele heeri</u>	7
<u>Owenia fusiformis</u>	7

Order Phyllodocida

Family Aphroditidae

<u>Aphrodita japonica</u>	5,7
<u>Aphrodita magellanica</u>	5
<u>Aphrodita refulgida</u>	7
<u>Heteaphrodita altoni</u>	5

Family Chrysopetalidae

<u>Paleanotus chrysolepsis</u>	7
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Family Glyceridae

<u>Glycera americana</u>	7
<u>Glycera capitata</u>	7

BENTHOS

<u>Glycera</u> <u>profundi</u>	7
<u>Glycera</u> <u>robusta</u>	7
<u>Glycera</u> <u>tenuis</u>	7
<u>Glycera</u> <u>tesselata</u>	5,7
Family Goniadidae	
<u>Bathyglycinde</u> <u>cedroensis</u>	7
<u>Glycinde</u> <u>armigera</u>	7
<u>Glycinde</u> <u>pacifica</u>	7
<u>Glycinde</u> <u>polygnatha</u>	7
<u>Gonaida</u> <u>annulata</u>	7
<u>Hemipodus</u> <u>borealis</u>	7
Family Hesionidae	
<u>Gyptis</u> <u>hyans</u>	7
<u>Podarke</u> <u>pugettensis</u>	7
<u>Podarkeopsis</u> <u>brevipalpa</u>	7
Family Nephtyidae	
<u>Aglaophamus</u> <u>eugeniae</u>	7
<u>Nephtys</u> <u>caeca</u>	7
<u>Nephtys</u> <u>caecoides</u>	7
<u>Nephtys</u> <u>californienesis</u>	7
<u>Nephtys</u> <u>ciliata</u>	7
<u>Nephtys</u> <u>cornuta</u>	7
<u>Nephtys</u> <u>ferruginea</u>	7
<u>Nephtys</u> <u>longosetosa</u>	7
<u>Nephtys</u> <u>punctata</u>	7
Family Nereidae	
<u>Ceratocephala</u> <u>loveni</u>	7
<u>Cheilonereis</u> <u>cyclurus</u>	7
<u>Neanthes</u> <u>brandti</u>	7
<u>Nereis</u> <u>eakini</u>	7
<u>Nereis</u> <u>mediator</u>	7
<u>Nereis</u> <u>neoneanthes</u>	7
<u>Nereis</u> <u>pelagica</u>	7
<u>Nereis</u> <u>pelagica</u> <u>neonigripes</u>	7
<u>Nereis</u> <u>procera</u>	7
<u>Nereis</u> <u>vexillosa</u>	7
<u>Nereis</u> <u>zonata</u>	7
<u>Platynereis</u> <u>bicanaliculata</u>	7
Family Pholoididae	
<u>Pholoides</u> <u>aspera</u>	7

BENTHOS

Family Phyllodocidae

<u>Anaitides groenlandica</u>	7
<u>Anaitides medipapillata</u>	7
<u>Anaitides williamsi</u>	7
<u>Austrophyllum exsilium</u>	7
<u>Chaetoparia careyi</u>	7
<u>Eteone pacifica</u>	7
<u>Eulalia bilineata</u>	7
<u>Eulalia quadrioculata</u>	7
<u>Eumida fusigera</u>	7
<u>Eumida sanguinea</u>	5,7
<u>Genetyllis castanea</u>	7
<u>Lugia abyssicola</u>	7
<u>Notophyllum imbricatum</u>	7
<u>Paranaitis polynoides</u>	7
<u>Protomystides Occidentalis</u>	7
<u>Pterocirrus imajimai</u>	7

Family Pilagiidae

<u>Ancistrostylis breviceps</u>	7
<u>Ancistrostylis groenlandica</u>	7
<u>Sigambra setosa</u>	7
<u>Sigambra tentaculata</u>	7
<u>Synelmis klatti</u>	7

Family Polynoidae

<u>Arctonoe fragilis</u>	7
<u>Arctonoe vittata</u>	7
<u>Eucranta anoculata</u>	7
<u>Eunoe nodosa</u>	7
<u>Halocydna brevisetosa</u>	7
<u>Harmathoe extenuata</u>	5
<u>Harmathoe imbricata</u>	7
<u>Harmathoe lunulata</u>	7
<u>Hesperonoe complanata</u>	7
<u>Hololepidella tuta</u>	7
<u>Lagisca multisetosa</u>	7
<u>Lepidonotus squamatus</u>	5,7
<u>Nemidia canadensis</u>	7

Family Pholoididae

<u>Pholoides aspera</u>	7
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Family Syllidae

<u>Autolytus prismaticus</u>	7
<u>Exogone gemmifera</u>	7
<u>Odontosyllis phosphorea</u>	7
<u>Sphaerosyllis californiensis</u>	7

BENTHOS

<u>Syllis elongata</u>	7
<u>Trypanosyllis adamanteus</u>	7
<u>Tryposyllis gemmipara</u>	7
<u>Tryposyllis armillaris</u>	7
<u>Tryposyllis hyalina</u>	7
<u>Tryposyllis pulchra</u>	7
Family Tomopteridae	
<u>Tomopteris septentrionalis</u>	7
Order Sabellida	
Family Sabellidae	
<u>Bathyadmetella commando</u>	5
<u>Chone gracilis</u>	7
<u>Chone mollis</u>	7
<u>Euchone analis</u>	7
<u>Eudistyllia vancouveri</u>	7
<u>Fabrisabella similis</u>	7
<u>Megalomma splendida</u>	7
<u>Potamethus mucronatus</u>	7
<u>Potamilla ocelata</u>	7
<u>Sabella media</u>	7
<u>Schzobrnachia insignis</u>	7
Family Serpulidae	
<u>Circeis spirillum</u>	7
<u>Serpul's avermicularis</u>	7
<u>Spirorbis borealis</u>	7
Order Spionidae	
Family Chaetopteridae	
<u>Chaetopterus variopedatus</u>	7
<u>Phyllochaetopterus limicolus</u>	7
Family Cirratulidae	
<u>Chaetozone corona</u>	7
<u>Chaetozone setosa</u>	7
<u>Cirratulus cirratus</u>	5,7
<u>Dirriformia spirabrancha</u>	7
<u>Dodecaceria fistulicola</u>	7
<u>Tharyx monilaris</u>	7
<u>Tharyx multifilis</u>	7
<u>Tharyx multifolis</u>	7

BENTHOS

Family Magelonidae

<u>Magelona</u> <u>cerae</u>	7
<u>Magelona</u> <u>japonica</u>	7
<u>Magelona</u> <u>papillicornis</u>	7
<u>Magelona</u> <u>pitelkae</u>	7

Family Spionidae

<u>Boccardia</u> <u>proboscidea</u>	7
<u>Laetmonice</u> <u>pellucida</u>	5
<u>Laonice</u> <u>cirrata</u>	7
<u>Minuspio</u> <u>cirrifer</u>	7
<u>Minuspio</u> <u>minor</u>	7
<u>Minuspio</u> <u>sp.A</u>	7
<u>Paraprionosio</u> <u>pinnata</u>	7
<u>Polydora</u> <u>armata</u>	7
<u>Polydora</u> <u>brachicephala</u>	7
<u>Polydora</u> <u>cardalia</u>	7
<u>Polydora</u> <u>commensalis</u>	7
<u>Polydora</u> <u>ligni</u>	7
<u>Polydora</u> <u>socialis</u>	7
<u>Prionospio</u> <u>anuncata</u>	7
<u>Prionospio</u> <u>malmgreni</u>	7
<u>Scoloepis</u> <u>foliosa</u>	7
<u>Scololepis</u> <u>squamatus</u>	7
<u>Spiophanella</u> <u>pallida</u>	7
<u>Spiophanes</u> <u>anoculata</u>	7
<u>Spiophanes</u> <u>berkeleyorum</u>	7
<u>Spiophanes</u> <u>bombyx</u>	6,7
<u>Spiophanes</u> <u>cirrata</u>	7
<u>Spiophanes</u> <u>fimbriata</u>	7
<u>Spiophanes</u> <u>kroeyeri</u>	7

Order Sternaspida

Family Stermaspidae

<u>Stermaspis</u> <u>fossor</u>	
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Order Terebellida

Family Ampharetidae

<u>Amage</u> <u>anops</u>	7
<u>Amelinna</u> <u>abyssalis</u>	7
<u>Ampharete</u> <u>acutifrons</u>	7
<u>Ampharete</u> <u>goesi</u>	7
<u>Amphicteis</u> <u>mucronata</u>	7
<u>Amphicteis</u> <u>scaphobranchiata</u>	7
<u>Amphisamytha</u> <u>bioculata</u>	7
<u>Anobothrus</u> <u>gracilis</u>	7
<u>Jugamphicteis</u> <u>paleata</u>	7

BENTHOS

<u>Lysippe annectens</u>	7
<u>Melinna cristata</u>	5,7
<u>Melinnampharete gracilis</u>	7
<u>Schistocomus hiltoni</u>	7
Family Pectinariidae	
<u>Cistenides brevicoma</u>	7
<u>Pectinaria californiensis</u>	6,7
Family Sabellariidae	
<u>Idanthyrus ornamentatus</u>	7
<u>Sabellaria cementarium</u>	5,7
<u>Sabellaria gracilis</u>	7
Family Terebellidae	
<u>Artacama coniferi</u>	7
<u>Eupolymnia cresentis</u>	7
<u>Loimia medusa</u>	7
<u>Neoamphitrite robusta</u>	7
<u>Pista cristata</u>	5,7
<u>Pista elongata</u>	7
<u>Pista fasciata</u>	7
<u>Pista fimbriata</u>	5
<u>Pista pacifica</u>	7
<u>Thelepus crispus</u>	7
<u>Thelepus setosus</u>	5,7
Family Tricobrachidae	
<u>Artacamella hancocki</u>	7
<u>Filibrachus roseus</u>	7
<u>Terebellides stroemii</u>	5,7
<u>Trichobranchus glacialis</u>	7
Phylum Pogonophora	
<u>Galathealinum brachiosum</u>	2
<u>Lamellasabella zachsi</u>	2
Order Vestimentifera	
<u>Lamellibrachia barhami</u>	3,4
Phylum Arthropoda	
Subphylum Chelicerata	
Class Pycnogonida	
<u>Colossendeis colossea</u>	5

BENTHOS

<u>Colossendeis angusta</u>	5
<u>Ascorhynchus japonicus</u> ?	5

Subphylum Crustacea

Class Cirripedia

<u>Amigdoscalpellum aurivillii incertum</u>	3,4
<u>Scalpellum albatrossianum</u>	2
<u>Scalpellum antillarum</u>	2
<u>Scalpellum larvale</u>	2
<u>Scalpellum perlongum</u>	2
<u>Scalpellum phantasma</u>	2
<u>Scalpellum regium</u>	2
<u>Scalpellum sanctaebarae</u>	2,5

Class Malacostraca

Order Mysidacea

<u>Ceratomyxis spinosa</u>	2
<u>Gnathophausia spp.</u>	5

Class Eumalacostraca

Order Amphipoda

<u>Acidostoma obesum</u>	9
<u>Amathillopsis pacifica</u>	9
<u>Ampelisca eoa</u>	9
<u>Ampelisca coeca</u>	9
<u>Ampelisca plumosa</u>	8,9
<u>Bathymedon caino</u>	9
<u>Bathymedon nepos</u>	9
<u>Bathymedon sp. 1</u>	8,9
<u>Bathymedon sp. 2</u>	8,9
<u>Bathymedon sp. 3</u>	9
<u>Bathymedon sp. 4</u>	9
<u>Bathymedon sp. 5</u>	9
<u>Bathymedon sp. 6</u>	9
<u>Bathymedon sp. a</u>	8,9,10
<u>Bathymedon sp. w</u>	9
<u>Bathymedon sp. z</u>	9
<u>Bonnierella linearis</u>	8,9
<u>Bruzelia inlex</u>	9
<u>Bruzelia sp. a</u>	9
<u>Byblis crassicornis</u>	8,9,10
<u>Corophid sp. a</u>	9
<u>Dulichia abyssii</u>	9
<u>Epimeria spp.</u>	9,10
<u>Gammarospsis spp.</u>	8,9
<u>Halice sp. a</u>	8,9
<u>Halice sp. w</u>	9

BENTHOS

<u>Halice sp. x</u>	9
<u>Halice sp. y</u>	9
<u>Halice sp. z</u>	8,9
<u>Haploops lodo</u>	9
<u>Haploops spp.</u>	10
<u>Harpiniopsis emeryi</u>	8,9,10
<u>Harpiniopsis excavata</u>	8,9,10
<u>Harpiniopsis fulgens</u>	8,9,10
<u>Harpiniopsis galerus</u>	10
<u>Harpiniopsis naiadis</u>	8,10
<u>Harpiniopsis percellaris</u>	8,10
<u>Harpiniopsis triplex</u>	8,9,10
<u>Harpiniopsis sp. 1</u>	8,9
<u>Hippomedon nubifer</u>	10
<u>Hippomedon strages</u>	9
<u>Hippomedon tracatrix</u>	9
<u>Hippomedon sp.b</u>	9
<u>Koroga megalops</u>	9
<u>Lepechinella echinata</u>	9
<u>Lepechinella turpis</u>	9
<u>Lepechinella sp. a</u>	9
<u>Lepechinella sp. b</u>	9
<u>Lepidepcreum sp. a</u>	9
<u>Leucothoe uschakoui</u>	9
<u>Liljeborgia cota</u>	8,9,10
<u>Monoculodes diversisexus</u>	9
<u>Monoculodes latissimanus</u>	8,9
<u>Monoculodes necopinus</u>	8,9
<u>Monoculodes recandesco</u>	8,9,10
<u>Monoculodes sp. 1</u>	9
<u>Monoculodes sp. x</u>	9
<u>Monoculodes sp. y</u>	8,9
<u>Monoculodes sp. z</u>	9
<u>Oediceroides abyssorum</u>	9
<u>Oediceroides trepedora</u>	8,9
<u>Oediceroides sp. y</u>	8,9
<u>Orchomene tabasco</u>	9
<u>Paracentromedon sp. a</u>	9
<u>Paraphoxus oculatus</u>	9,10
<u>Pardalisca sp. a</u>	9
<u>Pardaliscella symmetrica</u>	10
<u>Pardaliscella sp. a</u>	8,9
<u>Pardaliscid sp. g</u>	9
<u>Pardaliscoides tikal</u>	8,9
<u>Pardaliscoides sp. a</u>	8,9
<u>Pardaliscoides sp. b.</u>	9
<u>Paraliscopsis copal</u>	9
<u>Phipsiella minima</u>	9
<u>Photis kurilica</u>	8,9
<u>Proboloides tunda</u>	8,9
<u>Pseudotriron spp.</u>	10
<u>Rhachotropis lucifidor</u>	9
<u>Rhachotropis multisimis</u>	9

BENTHOS

<u>Rhachotropis sp. a</u>	9
<u>Rhachotropis sp. b</u>	9
<u>Rhachotropis sp. c</u>	9
<u>Syrrhoe oluta</u>	8,9
<u>Syrrhoe sp. b</u>	8,9
<u>Syrrhoites sp. a</u>	8,9
<u>Tryphosella sp. a</u>	8,9
<u>Uristes perspinus</u>	8,9
<u>Uristes spp.</u>	10
<u>Urotroae rotundifrons</u>	8,9
<u>Waldekia sp. a</u>	9

Order Decapoda

<u>Chionoecetes angulatus</u>	5
<u>Chionoecetes tanneri</u>	2,5,6
<u>Crangon abyssorum</u>	2,5
<u>Crangon franciscorum</u>	2,5
<u>Eualus biunguis</u>	5
<u>Hymnodora frontalis</u>	5
<u>Lithodes couesi</u>	2,5
<u>Munidopsis aries</u>	11
<u>Munidopsis bairdii</u>	11
<u>Munidopsis beringana</u>	11
<u>Munidopsis cascadia</u>	11
<u>Munidopsis ciliata</u>	11
<u>Munidopsis latirostris</u>	11
<u>Munidopsis quadrata</u>	2,5,11
<u>Munidopsis scabra</u>	5
<u>Munidopsis subsquamosa</u>	11
<u>Munidopsis tuftsi</u>	11
<u>Munidopsis verrilli</u>	2
<u>Munidopsis verrucosus *</u>	11
<u>Munidopsis yaquinaensis</u>	11
<u>Munidopsis spp.</u>	2
<u>Notostomas japonicus</u>	5
<u>Oregonia bifurcata</u>	5
<u>Pandalopsis ampla</u>	2
<u>Pandalus spp.</u>	2
<u>Paralomis multispina</u>	2,5
<u>Paralomis verrilli</u>	2,5
<u>Parapagurus mertensii</u>	2
<u>Parapagurus pilosimanus</u>	5
<u>Sergetes similis</u>	5,6

Order Isopoda

<u>Aega symmetrica</u>	6
<u>Leptochelia dubia</u>	6
<u>Leptognathia longiremis</u>	6
<u>Parantanais nanalmdensis</u>	6

BENTHOS

Phylum Mollusca

Class Amphineura

<u>Leptochiton mesogonus</u>	2
<u>Leptochiton spp.</u>	2

Class Bivalvia

<u>Calyptogena magnifica</u> ?	3,4
<u>Cuspidaria spp.</u>	2
<u>Halicardissa perplicata</u>	5,12
<u>Lyonsiella alaskana</u>	5,12
<u>Malletia faba</u>	5
<u>Myonera tillamookensis</u>	5,12
<u>Nucula carlottensis</u>	5
<u>Nucula cardara</u>	6
<u>Poromya leonina</u>	12
<u>Solemya agassizi</u>	2,5,6
<u>Solemya spp.</u>	3,4
<u>Tindaria gibbsii</u>	5

Class Cephalopoda

Order Octopoda

<u>Benthoctopus spp.</u>	5
<u>Graneledone spp.</u>	5
<u>Japetella heathi</u>	5
<u>Polypus sp. b</u>	2

Class Gastropoda

Order Prosobranchiata

<u>Admete californica</u>	5
<u>Antiplanes vinosa</u>	2,5,6
<u>Bathydoris spp.</u>	2
<u>Buccinum diplosetum</u>	2
<u>Buccinum viridum</u>	5
<u>Cidarina carlotta</u>	5
<u>Colus dimidiatus</u>	2,5
<u>Colus sapidus</u>	2
<u>Colus trophius</u>	5
<u>Leucosyrinx amycus</u>	2
<u>Leucosyrinx persimilis</u>	5
<u>Leucosyrinx persimilis blanca</u>	5
<u>Leucosyrinx persimilis leonis</u>	5
<u>Mohnia vernalis</u>	5
<u>Pleurotomella herminea</u>	2
<u>Plicifusus spp.</u>	2
<u>Solariella permeabilis</u>	2,5
<u>Tritonia diomeda</u>	2

BENTHOS

Class Scaphopoda

<u>Cadulus californicus</u>	2,6,13
<u>Dentalium agassizi</u>	2,13
<u>Dentalium ceras</u>	2
<u>Dentalium dalli</u>	5
<u>Dentalium megasthyris</u>	2,6
<u>Dentalium rectius</u>	6,13
<u>Dentalium sp. b</u>	2

Phylum Echinodermata

Class Asteroidea

<u>Asthenactis fisheri</u>	5,16
<u>Astrocles actinodotus</u>	2,10
<u>Astrolirus panamensis</u>	2
<u>Benthopecten acanthonotus</u>	2,5,16
<u>Benthopecten claviger</u>	2,5,6,16
<u>Benthopecten spp.</u>	2,10,14,15
<u>Brisingella exilis</u>	2,5,16
<u>Brisingia spp.</u>	2,16
<u>Ctenodiscus crispatus</u>	2,5,10,14,15,16
<u>Dipsacaster anoplus</u>	2,5,10,14,15,16
<u>Dytaster gilberti *</u>	1,2,10,14
<u>Dytaster spp.</u>	2
<u>Eremicaster pacificus</u>	2,5,10,14,15,16
<u>Eremicaster spp.</u>	2
<u>Heterozonias alternatus</u>	2,5,6,10,14,15,16
<u>Hippasteria californica</u>	2,5,6,10,14,15,16
<u>Hymenaster perissonotus</u>	5,16
<u>Hymenaster quadrispinosus</u>	2,5,6,10,14,15,16
<u>Hymenaster spp.</u>	2,5,10
<u>Leptychaster anomalous</u>	2,6
<u>Lophaster furciliger</u>	2,5,10,14,15,16
<u>Mediaster elegans</u>	14
<u>Mediaster elegans abyssi</u>	2,15
<u>Myxoderma sacculatum</u>	2,5,6,16
<u>Myxoderma spp.</u>	2
<u>Nearchaster asiculosus</u>	2,5,10,14,15,16
<u>Pectinaster agassizi evoplus</u>	2,5,16
<u>Pectinaster spp.</u>	14,15
<u>Pseudarchaster parelii alascensis</u>	2,5,10,15,16
<u>Pseudarchaster dissonus</u>	2,5,10,14,15,16
<u>Pseudarchaster spp.</u>	2,10
<u>Psilaster pectinatus</u>	2,5,6,10,14,15,16
<u>Pteraster coscinopeplus</u>	5,16
<u>Pteraster jordani</u>	2,5,16
<u>Solaster borealis</u>	2,5,6,10,14,15,16
<u>Zoroaster ophiurus</u>	2,5,15,16

BENTHOS

Class Crinoidea

<u>Florometra asperrima</u>	5,16
<u>Florometra serratissima</u>	5,16
<u>Psathyrometra fragilis</u>	5,16
Family Hyocrinidae *	1

Class Echinoidea

<u>Aeropsis fulva</u>	2,17
<u>Ceratophysa rosea</u>	2,17
<u>Sperosoma giganteum</u>	2,10,17
<u>Urechinus loveni</u>	2,17

Class Holothuroidea

<u>Abyssocucumis albatrossi</u>	2,10,18,19,20
<u>Amperima naresi</u>	19,20
<u>Amperima rosea</u>	19,20
<u>Amperima spp.</u>	5,16,18
<u>Benthodytes incerta</u>	19,20
<u>Benthodytes sanguinolenta</u>	18,19,20
<u>Caphiera sulcata</u>	18,19,20
<u>Ceraplectana trachyderma</u>	18
<u>Chirodota albatrossi</u>	2,10,18
<u>Laetmagone wyvillethomsoni</u>	18,19,20
<u>Laetmophasma fecundum</u>	2,6,10,18
<u>Laetmophasma spp.</u>	2
<u>Leptosynapta sp. b</u>	2
<u>Mesothuria murrayi</u>	19,20
<u>Molpadia granulata</u>	2,10,18,19,20
<u>Molpadia intermedia</u>	2,5,10,18
<u>Molpadia musculus</u>	2,5,10,16,18,19,20
<u>Molpadia spinosa</u>	2,10
<u>Molpadia spp.</u>	2,10
<u>Myriotrochus bathibius</u>	18
<u>Myriotrochus giganteus</u>	18
<u>Myriotrochus spp.</u>	18
<u>Onerophanta mutabilis</u>	18,19,20
<u>Paelopatides confundens</u>	5,18,19,20
<u>Paelopatides spp.</u>	2,6,10,16
<u>Pannychia moselyi</u>	2,5,10,16,18,19,20
<u>Peniagone dubia</u>	19,20
<u>Peniagone gracilis</u>	19,20
<u>Peniagone spp.</u>	2,10,18
<u>Protankyra duodactyla</u>	5,16,18
<u>Protankyra pacifica</u>	18
<u>Protankyra spp.</u>	5,16
<u>Pseudostichopus mollis</u>	5,16
<u>Pseudostichopus nudus</u>	2,10,18,19,20
<u>Pseudostichopus villosus</u>	19,20
<u>Pseudostichopus sp. a</u>	2

BENTHOS

<u>Pseudostichopus sp. b</u>	2
<u>Pseudostichopus sp. c</u>	2
<u>Pseudostichopus spp.</u>	10,18
<u>Psychropotes longicaudata</u>	19,20
<u>Psychropotes raripes</u>	2,10,18
<u>Scotoplanes clarki</u>	19,20
<u>Scotoplanes globosa</u>	2,6,18,19,20
<u>Scotoplanes theeli</u>	2,5,10,16,18
<u>Scotoplanes spp.</u>	2,10
<u>Synallactes gilberti</u>	2,10,18,19,20
<u>Synallactes spp.</u>	2,10
<u>Ypsilothuria bitentaculata</u>	5,16,19,20
<u>Zygothurea lactea</u>	16

Class Ophiuroidea

<u>Amphiacantha gastracantha</u>	21
<u>Amphilepsis platytata</u>	21
<u>Amphiophiura ponderosa</u>	5,21
<u>Amphiophiura superba</u>	21
<u>Amphiura diomedae</u>	2
<u>Amphiura korae</u>	5,16,21
<u>Amphiura spp.</u>	2
<u>Asteronyx loveni</u>	2,5,10,16,21
<u>Ophicantha bathybia</u>	5,21
<u>Ophiacantha normani</u>	2,10
<u>Ophiacantha trachybactra</u>	21
<u>Ophiacantha spp.</u>	2,10
<u>Ophiacanthella acontophora</u>	21
<u>Ophimitra spp.</u>	2,10
<u>Ophiocten pacificum</u>	2,5,10,16,21
<u>Ophiolimna bairdii</u>	5,16,21
<u>Ophiomusium jolliensis</u>	2,5,10,16,21
<u>Ophiomusium lymani</u>	2,5,16,21
<u>Ophiomusium multispinum</u>	2,5,16,21
<u>Ophiomusium spp.</u>	2
<u>Ophiophtalmus eurypoma</u>	21
<u>Ophiophtalmus normani</u>	5,16,21
<u>Ophiura bathybia</u>	21
<u>Ophiura hadra</u>	5,10,16,121
<u>Ophiura irrorata</u>	2,5,10,16,21
<u>Ophiura leptoctenia</u>	5,16,21
<u>Ophiura spp</u>	2,10
<u>Pandelia carchara</u>	21

BENTHOS

Appendix 2

Species found outside the lease area that may occur in the Gorda Ridge lease area, including hydrothermal vent organisms

MASTER REFERENCES:

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2. Ruby & Jannasch 1982
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7. Hessler & Smithey 1984
8. Burrenson 1981
9. Felbeck et al 1983
10. Desbruyeres & Laubier 1980
11. Banse 1979
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16. Pettibone 1984
17. Maciolek 1981
18. Jones 1980
19. Krantz 1982
20. Newman 1979
21. Smith 1984
22. Humes & Dojiri 1980
23. Carey & Alspach 1966
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27. Williams & Chase 1982
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29. Williams & Van Dover 1983
30. Hessler 1984
31. Boss & Turner 1980
32. Bernard 1974
33. Alton 1972
34. Carey 1963
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36. Carey et al 1967
37. Jannasch 1984b

Bacteria

Reference no.

<u>Beggiatoa</u>	37
<u>Hyphomonas spp.</u>	37
<u>Methanococcus jannaschii</u>	1
<u>Thiobacillus spp.</u>	37
<u>Thiomicrospira spp.</u> Strain L.-12	2

BENTHOS

Phylum Porifera

Class Hexactinellida

Phylum Coelenterata

Class Anthozoa

<u>Anthoptilum grandiflorum</u>	4
<u>Actinaria spp.</u>	5
<u>Balticina pacifica</u>	4
<u>Paraphelliactis pabista</u>	6
<u>Pennatulacea spp.</u>	5

Phylum Enteropneusta

Phylum Annelida

Class Hirudinea

<u>Bathybdella sawyeri</u>	8
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Class Oligochaeta

<u>Phallodrilus leukodermata</u>	9
<u>Phallodrilus planus</u>	9

Class Polychaeta

<u>Alvinella pompejana</u>	10
<u>Ampharetidae spp.</u>	11
<u>Amphisamytha galapagensis</u>	12
<u>Eunicidae spp.</u>	13
<u>Harmathoe tenebricosa</u>	3
<u>Paralvinella grasslei</u>	14
<u>Polynoidae spp.</u>	15,16
<u>Spionidae spp.</u>	17
<u>Serpulidae spp.</u>	7

Phylum Pogonophora

<u>Riftia pachyptila</u>	18
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Phylum Arthropoda

Subphylum Chelicerata

Class Acarina

<u>Copidognathus spp.</u>	19
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Subphylum Crustacea

BENTHOS

Class Cirripedia

<u>Scalpellidae</u> spp.	20
<u>Scalpellum</u> sp. b	4
<u>Lepas</u> spp.	4

Class Copepoda

<u>Isaacsicalanus paucisetus</u>	21
<u>Siphonostomid</u> spp.	22

Class Eumalacostraca

Order Amphipoda

<u>Byblis barbarensis</u>	23
<u>Byblis</u> spp.	24
<u>Europisa elongatum</u>	23
<u>Eurythenes gryllus</u>	25, 26
<u>Haploops</u> spp.	24
<u>Orchomene gerulicorbis</u>	25
<u>Paraliscella caperesca</u>	25
<u>Paraliscella tenuipes</u>	25
<u>Pardaliscid</u> spp.	23

Order Decapoda

<u>Acanthophyra</u> spp.	3
<u>Bresilidae</u> spp.	27
<u>Bythograea thermydron</u>	28
<u>Calastacus</u> spp.	3
<u>Chlorilia longipes turgida</u>	3
<u>Gennadas</u> spp.	3
<u>Munidopsis lentigo</u>	21
<u>Munidopsis</u> spp.	29
<u>Munida quadraspina</u>	3
<u>Nematocarcinus</u> spp.	7
<u>Pagurus capillatus</u>	4
<u>Pagurus tanneri</u>	3
<u>Spirontocaris flexa</u>	4

Order Leptostraca

<u>Dahlella calderensis</u>	30
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Phylum Mollusca

Class Bivalvia

<u>Calyptogena magnifica</u>	31
<u>Cardiomya planetica</u>	3
<u>Cuspidaria apodema</u>	32
<u>Cyclopecten randolphi</u>	3, 4

BENTHOS

<u>Delectopecten vancouverensis</u>	3
<u>Mytilidae spp.</u>	7
<u>Poromya malespinae</u>	32
<u>Xylophaga washingtona</u>	3
<u>Yoldia limatula gairdneri</u>	4
<u>Yoldia montereyensis</u>	3

Class Gastropoda

<u>Ancistrolepis spp.</u>	3
<u>Buccinum strigillatum</u>	3
<u>Buccinum tenue</u>	4
<u>Buccinum spp.</u>	4
<u>Colus aphelus</u>	3
<u>Colus halidonus</u>	3
<u>Neomphalus fretterae</u>	33
<u>Neptuna amianta</u>	3
<u>Plicifusus griseus</u>	3
<u>Tritonia spp.</u>	4
<u>Turidae spp.</u>	7

Phylum Echinodermata

Class Asteroidea

<u>Ampheraster marianus</u>	3, 4, 23
<u>Rathbunaster spp.</u>	4
<u>Thrissacanthias penicillatus</u>	3, 4, 23
	34, 35, 36, 37
<u>Zoraster evermani mordax</u>	3, 4, 23, 34, 36

Class Holothuroidea

<u>Molpadia oolitica</u>	4
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Class Ophiuroidea

<u>Ophiuroidae spp.</u>	7
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